# **Clostridia: Microbial Engines of Bioconversion**

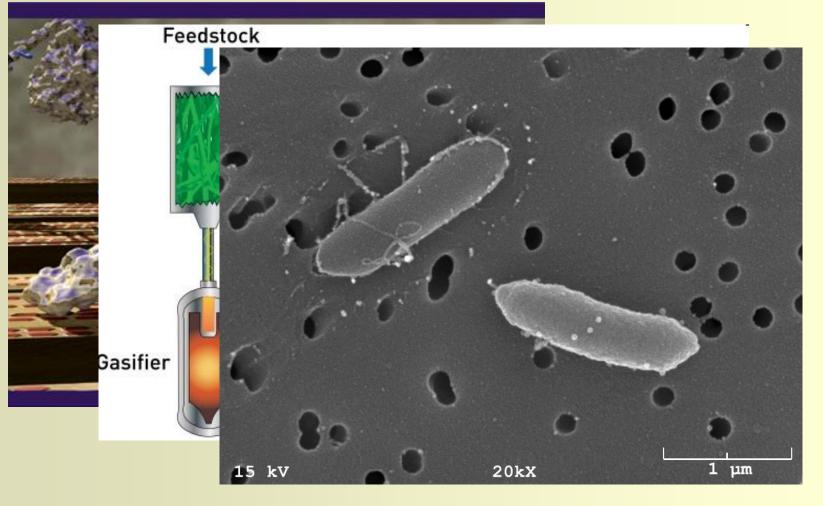
#### Oklahoma Annual EPSCoR State Conference April 29, 2010

**Chris Hemme and Jizhong Zhou** 



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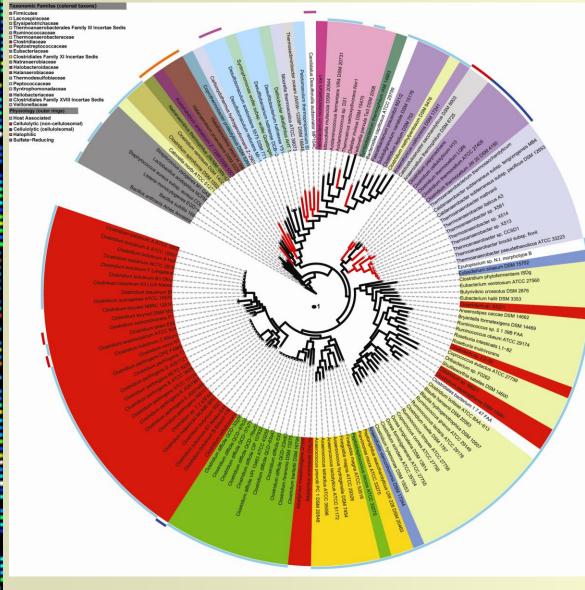
### **Clostridia: Microbial Engines of Bioconversion**





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## **Clostridia Phylogenomics**

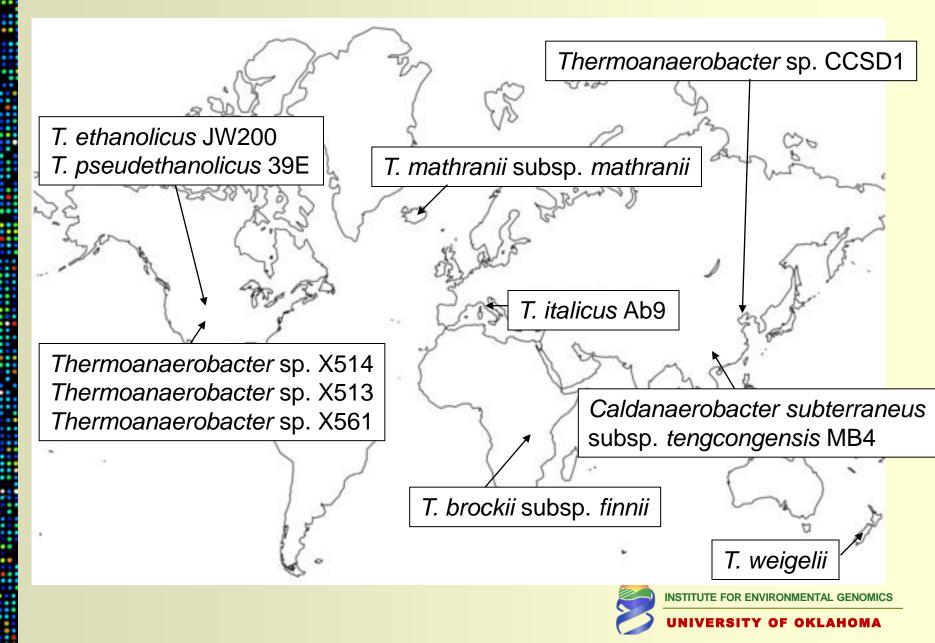


- Taxa colored based on 16S rRNA classification
- Red lineages = thermophiles
- Suggests taxonomic relationship between thermophilic saccharophiles and cellulosomeencoding Clostridia



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#### **Thermoanaerobacter** Genomics



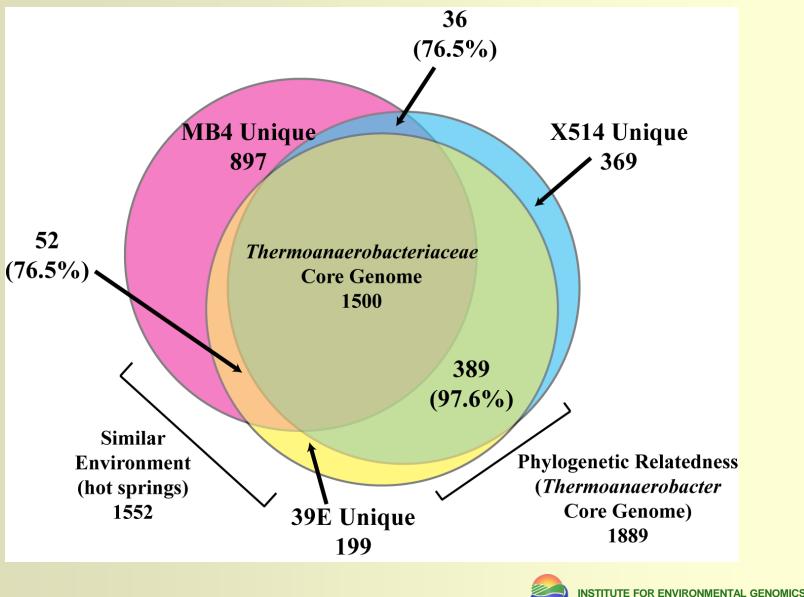
#### **Thermoanaerobacter Genomics**

- Comparative genomics
- Correlation of genomics to physiology
- Development of genetic systems



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#### **Thermoanaerobacter** Core Genome



#### **Results**

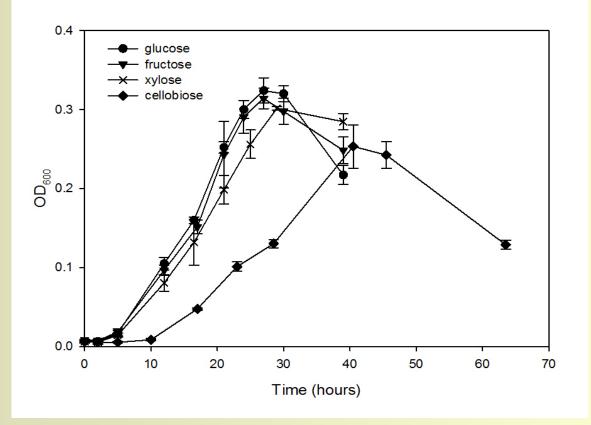
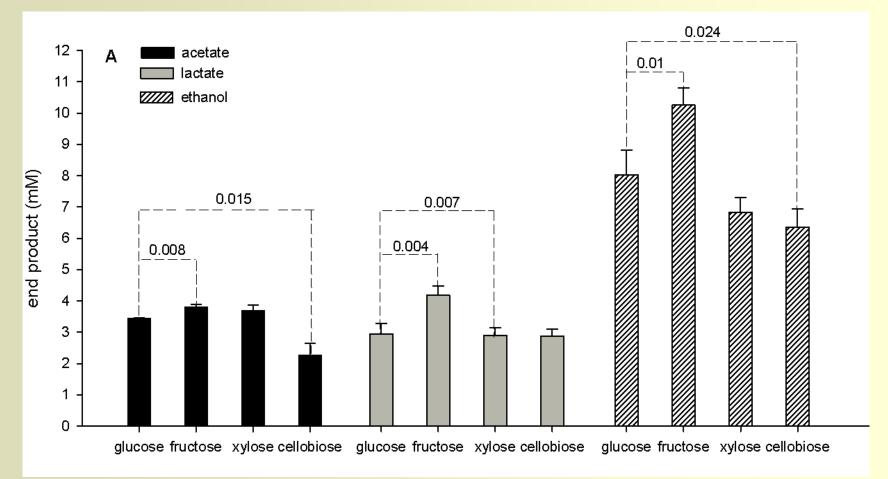


Fig. 1 Growth curves of Thermoanaerobacter sp. X514 grown in defined original medium with glucose, xylose, fructose and cellobiose as the sole carbon source. INSTITUTE FOR ENVIRONMENTAL GENOMICS



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## **Results**



**Fig. 2A Metabolite levels of** *Thermoanaerobacter* **sp. X514.** Metabolite production by *Thermoanaerobacter* **sp. X514** in defined medium under different substrate conditions.

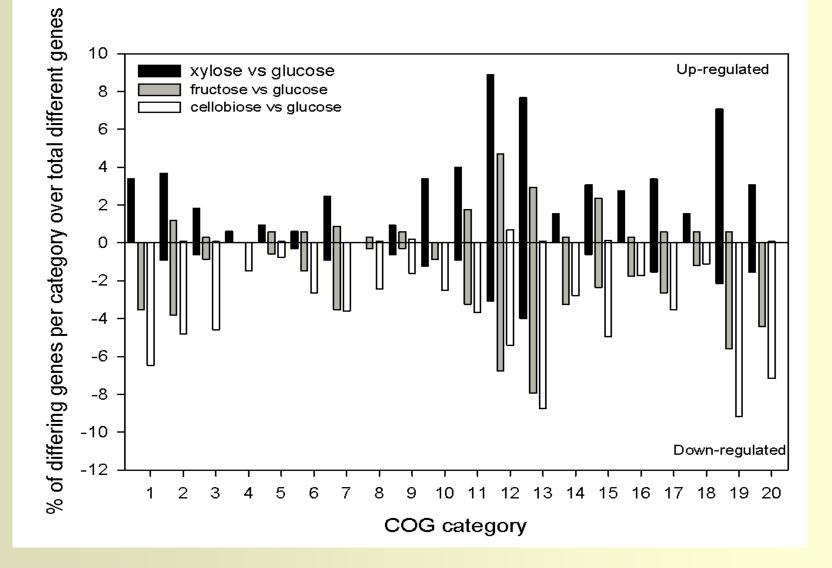
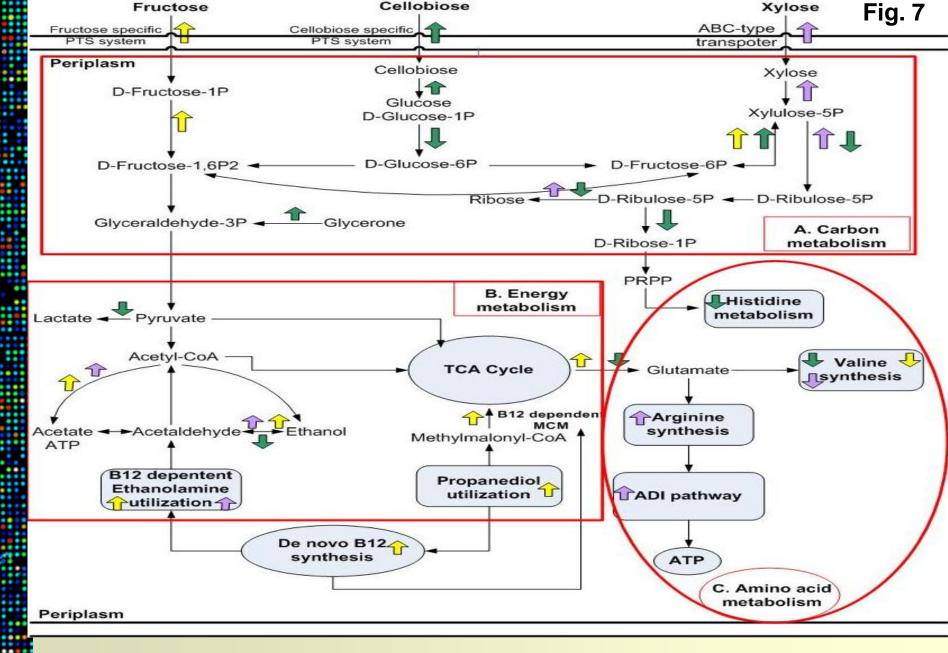
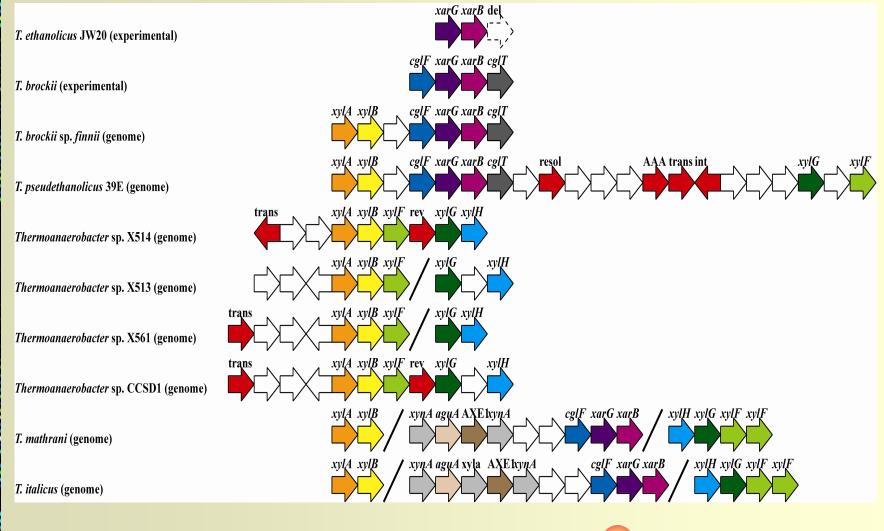


Fig. 3 Number of differentially expressed genes represented as a percentage of the total genes per functional classification according to JGI. Columns: 1, J; 2 K; 3 L; 4 D; 5 V; 6 T; 7 M; 8 N; 9 U; 10 O; 11 C; 12 G; 13 E; 14 F; 15 H; 16 I; 17 P; 18 Q; 19 Rin 20 Sitty of OKLAHOMA



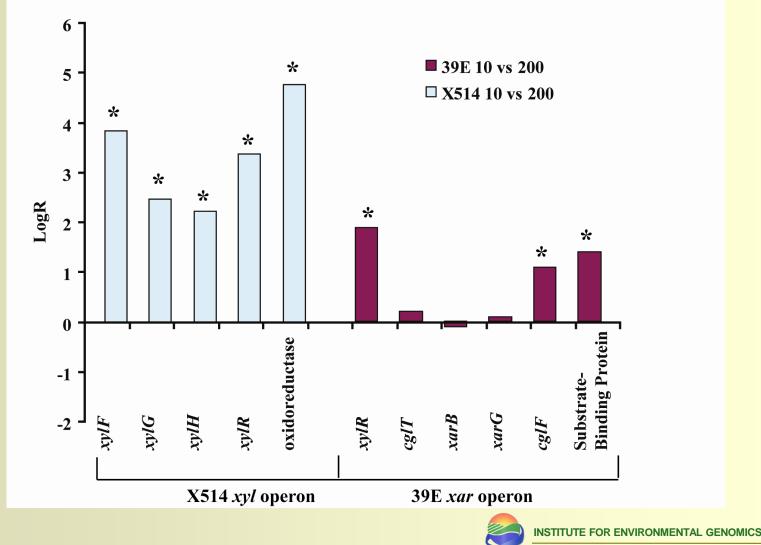
Notes: up arrow: up-regulation, down arrow: down-regulation, up-regulation, genomics yellow: fructose, green: cellobiose, purple: xylose

#### **Xylose and Xylan Metabolism and Transport in** *Thermoanaerobacter*

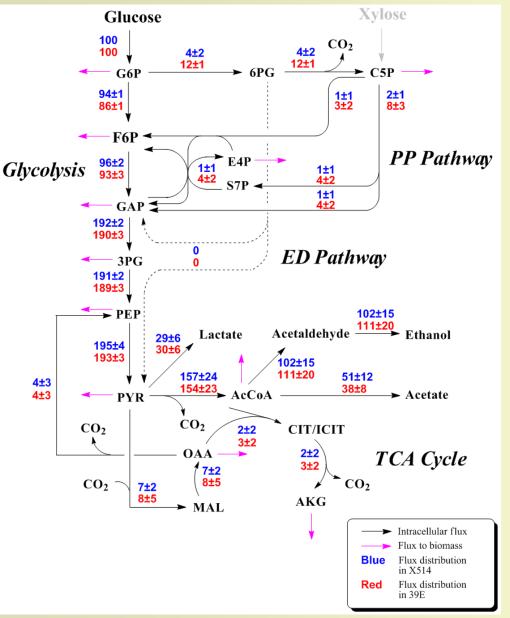




#### **Expression of Xylose Transport Genes**



### **Metabolic Flux Analysis**



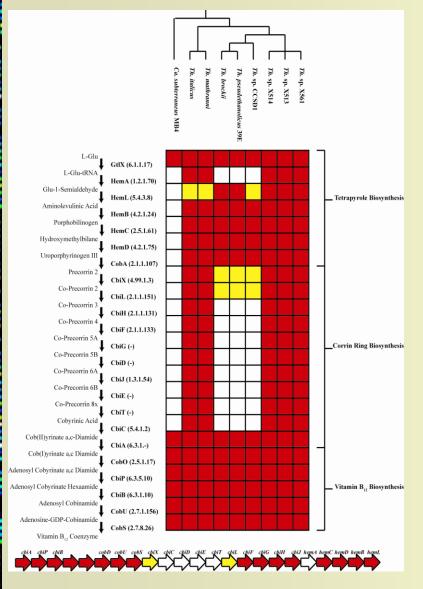
- Relative carbon flux from xylose the same in both strains
- Absolute carbon flux from xylose greater in X514



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#### Vitamin B<sub>12</sub> Biosynthesis in *Thermoanaerobacter*



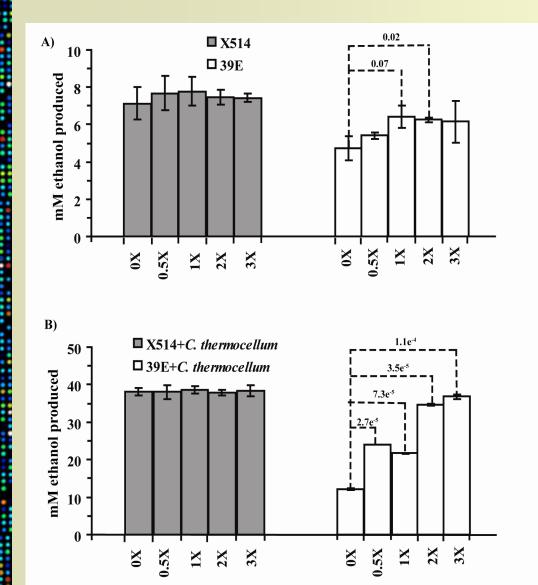
#### Complete *de novo* vitamin B<sub>12</sub> biosynthesis pathway in X514 and mathranii lineages

- Hypothesis
  - B<sub>12</sub> produced by X514 enhances cellulose degradation and ethanol yields in coculture w/ LQRI
- Experiments
  - Growth assays w/ LQRI monoculture grown on cellulose w/ B<sub>12</sub>
  - Growth assays w/ 39E-LQRI coculture grown on cellulose w/ B<sub>12</sub>



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## **Effects of Vitamin B<sub>12</sub> on Ethanol Production in Mono- and Co-Culture**



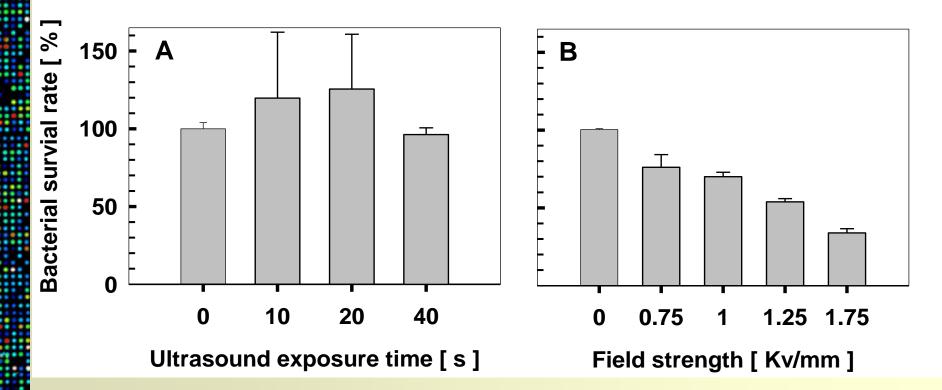
- X514 is insensitive to exogenous B<sub>12</sub>
- 39E is very sensitive to exogenous B<sub>12</sub>



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# **Results**

#### Survival rate of *Thermoanaerobacter* sp. X514 after sonoporation and electroporation.



The result indicated ultrasound can be used to efficiently incorporate macromolecules into X514 without significant cell damage.



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### **Results**

#### Effect of ultrasound exposure times on plasmid integrity

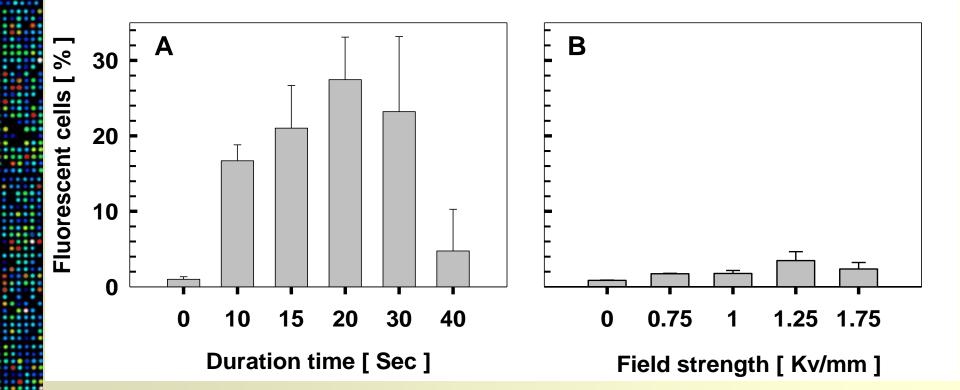
ultrasound exposure time [s] 10 15 20 30 40 0 M kb 10 8.0 6.0 5.0 4.0 3.0 2.0 1.5 1.2 1.0 **n\_9** 8 07 0.6 0.5n 4 0.3 0.2 0.1

The plasmid pIKM1 was separated on 1% agarose gel after ultrasound exposure with times ranged from 0 to 40 s in Branson B200 sonifier.

The result showed plasmid integrity was not significantly impacted by sonification at the exposure times investigated.



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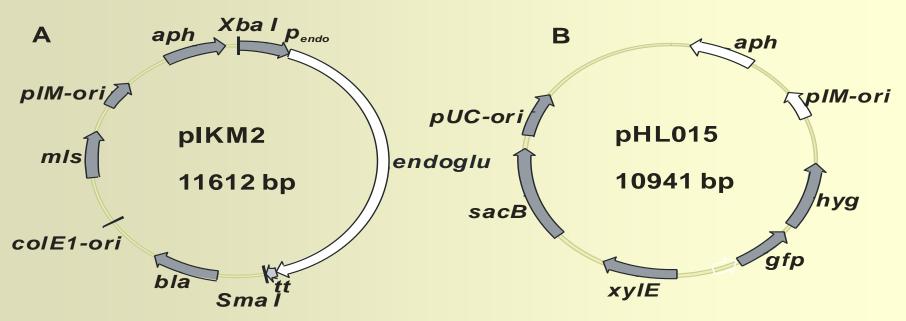


The optimal sonoporation parameters: 40kHz,19W,20S (Branson 200)

**Results** 



# Transformation of methylated pIKM2 and pHL015 into *Thermoanaerobacter* sp. X514 by sonoporation and electroporation



Physical maps of plasmid plKM2 and pHL015.

A: pIKM2. pIKM2 is a derivate of pIKM1 (22) by inserting the beta-1, 4-endoglucanase gene into *Xbal / Smal* sites of pIKM1 as described in text.

**B: pHL015.** pHL015 is a derivate of construct by inserting the *Ncol / Nsi* flanked fragment of pIKM1 into similarly digested plasmid pML523.

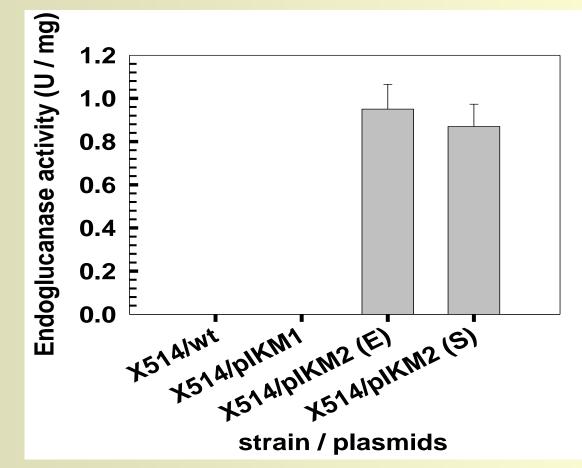
The pHL015 contains a kanamycin resistance gene (aph, from pIKM1),

a Bacillus subtilis replicon (pIM-ori, from pIKM1), a hygromycin resistance gene (hyg),

a green fluorescence protein gene (gfp), a catechol 2,3-dioxygenase gene (xy/E),

a levansucrase gene (sacB) and an E. coli replicon (pUC-ori)

# Endoglucanase activity of *Thermoanaerobter sp.* X514 transfomants



The X514 was transformed by electroporation (E) and sonoporation (S), respectively. Wild-type X514 and pIKM1 transformants were used as negative control.



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## **Future Directions**

- Establishment and refinement of genetic and mutagenic systems for Clostridia
- Long-term evolution of Thermoanaerobacter
- H<sub>2</sub> production from biomass
- More genomic sequencing and comparative genomics



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